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Abstract: **PURPOSE OF REVIEW** The intestinal immune system is constantly exposed to foreign antigens, which for the most part should be tolerated, but the immune system retains the ability to react rapidly and effectively to eliminate pathogens. Dendritic cells are at the front line in maintaining intestinal integrity as they are widely distributed within the intestinal lamina propria, Peyer's patches and mesenteric lymph nodes. **RECENT FINDINGS** The identification of dendritic cell subsets and phenotypic markers within the healthy and diseased intestine has progressed significantly, including improved identification of dendritic cell subsets within the human intestine. Recently, the role for dietary factors and the microbiome in modulating the intestinal dendritic cell functions has begun to be better investigated, resulting in a number of new findings relating to retinoic acid metabolism, pattern recognition receptor triggering and G-protein-coupled receptor activation. In addition, the interactions between goblet cells and mucin with intestinal dendritic cells are being better defined. **SUMMARY** In this review, we discuss the recent findings relating to intestinal dendritic cells, in particular the importance of dendritic cells in sensing the intestinal microenvironment and the consequences for health and disease.

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Intestinal dendritic cells

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Purpose of review

The intestinal immune system is constantly exposed to foreign antigens, which for the most part should be tolerated, but the immune system retains the ability to react rapidly and effectively to eliminate pathogens. Dendritic cells are at the front line in maintaining intestinal integrity as they are widely distributed within the intestinal lamina propria, Peyer's patches and mesenteric lymph nodes.

Recent findings

The identification of dendritic cell subsets and phenotypic markers within the healthy and diseased intestine has progressed significantly, including improved identification of dendritic cell subsets within the human intestine. Recently, the role for dietary factors and the microbiome in modulating the intestinal dendritic cell functions has begun to be better investigated, resulting in a number of new findings relating to retinoic acid metabolism, pattern recognition receptor triggering and G-protein-coupled receptor activation. In addition, the interactions between goblet cells and mucin with intestinal dendritic cells are being better defined.

Summary

In this review, we discuss the recent findings relating to intestinal dendritic cells, in particular the importance of dendritic cells in sensing the intestinal microenvironment and the consequences for health and disease.

Keywords

dendritic cells, goblet cells, microbiome, retinoic acid, short-chain fatty acids

INTRODUCTION

The intestine is a highly evolved organ specialized to perform the essential functions of nutrient digestion, absorption and waste disposal. The intestinal immune system has the momentous and unenviable task of maintaining intestinal integrity in the presence of an enormous quantity of external or foreign antigens. Highly sophisticated cellular and molecular networks need to be constantly coordinated in order to respond appropriately to such antigens, while also protective immune responses to potential pathogens must be maintained and can be induced effectively on demand. Inappropriate immune response to these antigens is a significant component in several intestinal diseases, including inflammatory bowel disease and food allergies. Dendritic cells are professional antigen-presenting cells that are present within all tissues exposed to the outside world as immature sentinel cells that efficiently sample their environment for foreign antigens [1,2]. In addition to their essential role in antigen presentation, dendritic cells express multiple pattern recognition receptors, which modulate their maturation resulting in the development of different dendritic cell subsets that selectively promote polarized lymphocyte

responses. Upon activation, dendritic cells undergo maturation into potent T-cell stimulatory effector or regulatory dendritic cells and migrate toward the T cell areas of draining lymphoid organs. There, dendritic cells will activate naive T_H cells with antigen-specific [major histocompatibility complex (MHC) peptide complexes, signal 1] and costimulatory or inhibitory (signal 2) molecules. Recently, CD83 expression by intestinal dendritic cells was shown to regulate dendritic cell activation and promote mucosal homeostasis [3[¶]]. In addition to signals 1 and 2, dendritic cells carry a third signal, which determines the polarization of naive T_H cells into T_H1, T_H2, T_H9, T_H17 or T regulatory cells [4]. Like signal 2, signal 3 is heterogeneous and can be mediated by various soluble or membrane-bound

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KEY POINTS

- Intestinal dendritic cells are key players in mediating mucosal regulatory responses, effector responses and pathological responses.
- The intestinal environment, including the microbiome and dietary factors, shape and condition intestinal dendritic cell activity.
- Goblet cells and mucins are involved in intestinal dendritic cell antigen sampling and modulation of dendritic cell activity.

molecules, including interleukin (IL)-1 β , IL-6, IL-10, IL-12, IL-18, TGF- β , IFN- α , OX40 ligand (OX40L) and retinoic acid [5,6]. Importantly, in-vitro studies suggest that the expression levels of these T_H-cell-polarizing molecules by mature dendritic cells strongly depend on the conditions during their initial activation as sentinel dendritic cells. These findings imply that pathogens, commensals and dietary components may promote the development of distinct dendritic cell phenotypes by provoking tissues to release mediators involved in polarization. The interplay between immunostimulatory and suppressive activities of dendritic cells is important for both the induction of an immune response and the maintenance of local immunostasis. The recent findings regarding the environmental factors influencing dendritic cell polarization will be discussed in the following review.

DENDRITIC CELL SUBSETS WITHIN THE INTESTINE

Intestinal dendritic cells are located within specific intestinal lymphoid tissues, collectively termed gut-associated lymphoid tissues (GALT), or diffusely distributed throughout the intestinal lamina propria [7]. Dendritic cell subsets are not evenly distributed along the intestinal tract, rather the CD103⁺CD11b⁺ dendritic cells are the major dendritic cell subset in the murine small intestine lamina propria, whereas in the colon these cells are reduced, resulting in higher proportions of CD103⁺CD11b⁻ and CD103⁻CD11b⁺ dendritic cells [8]. Dendritic cell subsets in the intestine have been extensively studied in mice, but studies in humans have been complicated by the realization that murine markers do not always correlate with human markers. Recently, a comparison between human and murine gut dendritic cell populations revealed that human CD103⁺ signal regulatory protein alpha (SIRP α)⁻ intestinal dendritic cells were related to murine intestinal CD103⁺CD11b⁻ dendritic cells. Human

CD103⁺SIRP α ⁺ dendritic cells are closely related to murine intestinal CD103⁺CD11b⁺ dendritic cells and they supported the induction of regulatory T cells. Both human CD103⁺ dendritic cell subsets induced T_H17 polarization, whereas CD103⁻SIRP α ⁺ dendritic cells promoted T_H1 polarization [9[■]]. These authors also performed comparative transcriptomics, which identified gene clusters that are evolutionarily conserved or divergent between mice and humans. The importance of SIRP α in the homeostasis of CD103⁺CD11b⁺ dendritic cells in the intestine was recently demonstrated, in which loss of SIRP α signaling in mice leads to a selective reduction in the CD103⁺CD11b⁺ dendritic cells within the mucosa associated with reduced numbers of T_H17 cells in steady-state intestinal mucosa and a defective T_H17 response to *Citrobacter rodentium* infection [10[■]]. The CD103⁻ dendritic cell subset has been associated with colitis severity and osteopontin expression by CD103⁻ dendritic cells drives intestinal inflammation. Disrupting the interaction between osteopontin and integrin α 9 suppressed the inflammatory potential of CD103⁻ dendritic cells *in vitro* and *in vivo* [11[■]].

DENDRITIC CELL MODULATION BY MICROBES AND MICROBIAL METABOLITES

The balance between immune tolerance and inflammation is regulated through the crosstalk between innate immune cells and the intestinal microbiota involving many signaling pathways and molecules. One component of these regulatory activities includes host-microbe-metabolic interactions and involves many organs, including the gut [12]. Intestinal dendritic cells are one of the main targets for immunomodulatory processes related to commensal microbes [13]. Metabolism of vitamin A to retinoic acid is a key immunomodulatory activity associated with intestinal dendritic cells [14[■]]. Certain, but not all, commensal microbes can induce retinoic acid metabolism by human dendritic cells *in vitro* and by murine CD103⁺ dendritic cells within the small intestine lamina propria [15[■],16]. Retinoic acid has also recently been described to play a role in immune regulation within the gastric mucosa. Gastric epithelial cells were described to promote retinoic acid metabolism in dendritic cells from the gastric mucosa. *Helicobacter pylori* infection severely disrupted gastric retinoic acid biosynthesis, which may lead to reduced dendritic cell retinoic acid signaling and may contribute to disease progression [17[■]]. In addition to vitamin A metabolism, induction of another dendritic cell metabolic enzyme, heme oxygenase-1 (HO-1), was shown to be required for the

induction of mucosal T regulatory cells within mesenteric lymph nodes by *Lactobacillus rhamnosus*. Upregulation of dendritic cell HO-1 resulted from direct contact with *L. rhamnosus* [18].

Bacterial cell wall components and metabolites [including cell-surface-associated or secreted polysaccharides, short-chain fatty acids (SCFAs), vitamins and biogenic amines, e.g., histamine] have been associated with immunoregulatory effects on dendritic cells at mucosal interfaces [19]. There is also evidence that these products can shape the immune response not only locally but also at extra-intestinal sites [20[■]], and they can act not only on dendritic cells but also directly on T cells, macrophages, invariant natural killer T cells and epithelial cells [21–23]. However, dendritic cells are the primary cell type involved as ‘sensors’ of microbial ligands through the activation of innate immune receptors (e.g. Toll-like receptors and c-type lectin receptors) expressed on their cell surface or inside the cytoplasm. The signaling pathways triggered by commensal-derived molecules allow for changes in dendritic cell phenotypes and cytokine secretion, which underlie the integration of microbial and host metabolism with immune functions.

Structural components of commensal microbes have been shown to be important for dendritic cell sensing of luminal bacterial species. For example, MHC-II-dependent antigen presentation of segmented filamentous bacteria antigens by intestinal CD11c⁺ dendritic cells is crucial for the local induction of T_H17 lymphocytes [24[■]]. In addition, a variety of polysaccharides from bacterial cell walls have been described to influence dendritic cell function. Capsular polysaccharide A (PSA) from the gut bacterium *Bacteroides fragilis* has been associated with regulatory properties. Its mechanism of action is not yet fully clear, but this polysaccharide has been shown to interact directly with mouse plasmacytoid dendritic cells via TLR-2. PSA-exposed plasmacytoid dendritic cells express molecules involved in protection against colitis and stimulated CD4⁺ cells to secrete IL-10 [25[■]]. In a multiple sclerosis mouse model, the same ligand protected against central nervous system demyelination and inflammation via TLR-2, and one of the possible mechanisms underlying these effects could involve dendritic cells [26]. PSA also interacts with human dendritic cell-specific Intercellular adhesion molecule-3-grabbing non-integrin (DC-SIGN) receptor expressed on monocyte-derived dendritic cells, which endocytose PSA thus facilitating presentation to T cells and the proliferation of PSA-specific CD4⁺ lymphocytes [27[■]]. Exopolysaccharides (EPS) which can be secreted or associated with the bacteria cell wall have also received attention for their potential

tolerogenic properties. EPS from *Bacillus subtilis* is able to prevent gut inflammation stimulated by *C. rodentium*, which is dependent on TLR-4 and MyD88 signaling in mouse myeloid cells including macrophages and dendritic cells [28[■]]. β-Glucans, which are polysaccharides found in bacteria, fungi and yeasts, are molecules which can have proinflammatory and anti-inflammatory properties and they have been studied for their potential benefits in cancer treatment and infection control. β-Glucan from *Saccharomyces cerevisiae* triggers murine dendritic cells to secrete TNF-α, IL-10 and TGF-β. The production of these cytokines as well as stimulation of indoleamine 2,3-dioxygenase (IDO) enzyme expression is dependent on Dectin-1 receptor engagement [29]. A population of Langerin-positive dendritic cells in murine Peyer’s patches was demonstrated to be involved in sampling β-glucan microparticles following transepithelial transport via M cells [30[■]]. Certain carbohydrate structures can have dual effects on dendritic cells and one example is the mannose-capped lipoarabinomannan (Man-LAM) from *Mycobacterium tuberculosis*. This lipoglycan has the ability to induce both proinflammatory (TNF, IL-6 and IL-12p40) and anti-inflammatory (IL-10) cytokines by dendritic cells in a Dectin-2-dependent manner [31]. Lastly, dietary-derived oligosaccharides can directly influence mucosal dendritic cells. The breast milk oligosaccharide sialyl(α2,3)lactose directly stimulated mesenteric lymph node CD11c⁺ dendritic cells to secrete cytokines required for the expansion of T_H1 and T_H17 T cells. The stimulatory effect was attenuated in Tlr4-deficient CD11c⁺ cells, demonstrating that this oligosaccharide induces inflammation through TLR-4 signaling [32[■]].

Recently, significant attention has been focused on the immunoregulatory role for SCFAs produced by commensal microbes within the gut. Species of *Roseburia*, *Eubacterium*, *Bacteroides* and *Faecalibacterium* are examples of bacteria that produce these metabolites in the gut ecosystem. The production of SCFAs occurs in the colon following fermentation of dietary fibers [33]. Abnormalities in the production of these metabolites (because of dietary factors and dysbiosis) might play a role in the pathogenesis of type 2 diabetes, obesity, inflammatory bowel disease, colorectal cancer and allergies [34]. Among the SCFAs, butyrate seems to be more potent than acetate or propionate in inducing immunomodulatory effects within the gut. Butyrate influences histone deacetylases (HDAC) activity, which is responsible for decreasing dendritic cell proinflammatory cytokine production (IL-12 and IL-6) and allows dendritic cells to promote T regulatory cells. Propionate is less potent than butyrate, although

propionate can also contribute to the induction of Foxp3 expression by dendritic cells, whereas acetate does not have this activity possibly because of the lack of HDAC activity [35[■]]. HDAC inhibition by butyrate has been also shown for intestinal macrophages [36]. Another recent study has confirmed and extended the observation that butyrate promotes dendritic cell regulatory activity, resulting in the induction of T regulatory cells and IL-10-producing T cells. These effects were mediated by the G-protein coupled receptor Gpr109a on colonic dendritic cells and macrophages, whereas Gpr109a-deficient mice were susceptible to the development of colonic inflammation and colon cancer [37[■]].

Histamine is another important mucosal metabolite and gut histamine levels are increased in patients with irritable bowel syndrome and inflammatory bowel disease [38[■]]. Histamine is able to decrease chemokine and proinflammatory cytokine secretion induced by Toll-like receptor stimulated dendritic cells, while increasing IL-10 production [39[■]]. Histamine exerted this effect by activating the histamine 2 receptor (H₂R) on dendritic cells, and the signaling mechanism required cAMP and exchange protein directly activated by cAMP (EPAC). Histamine is secreted not only by the host immune cells, but also by the bacteria within the gut. Administration of a histamine-secreting *Lactobacillus* strain to mice resulted in rapid weight loss and enhanced Peyer's patch cytokine secretion, which was exaggerated in H₂R-deficient animals [40[■]].

DENDRITIC CELL INTERACTIONS WITH GOBLET CELLS AND MUCINS

The gastrointestinal tract is covered by a mucus layer that has different properties in the stomach, small intestine and colon. In the small intestine, mucus limits the number of bacteria that can reach the epithelium and the Peyer's patches. In the large intestine, the inner mucus layer separates the commensal bacteria from the host epithelium. The outer colonic mucus layer is the natural habitat for the commensal bacteria. Mucus is secreted by goblet cells and typically contains several major components. One of these, the mucins gives it the mucus gel-like properties. The mucins produced by goblet cells are the classical gel-forming mucins MUC2, MUC5AC, MUC6 and MUC5B, whereas the transmembrane mucins MUC1, MUC3, MUC4, MUC12, MUC13 and MUC17 are all found in the gastrointestinal tract [41].

Previously, the function of goblet cells was thought to fulfill a purely secretory role within the intestine. However, recent evidence suggests that goblet cells in the small intestine deliver luminal

antigens to the dendritic cells within the lamina propria [42]. The uptake of luminal material by goblet cells is stimulated by acetyl choline analogs acting on muscarinic acetyl choline receptor 4 [43[■]]. Not all dendritic cell subsets sampled antigen from goblet cells, the CD11c⁺CD103⁺ dendritic cell subpopulation seems to be uniquely capable of performing this role. These dendritic cells can also cross-present antigens to T cells and induce tolerance, as described above. However, the mechanisms underpinning the goblet cells and CD103⁺ dendritic cells interactions that cooperate to promote tolerance is far from being understood, but it does seem that goblet cells have a new type of gate-keeping role for the presentation of oral antigen to dendritic cells within the lamina propria.

Mucin secreted by the goblet cells has also been recently shown to influence intestinal dendritic cell activity [44[■]]. Dendritic cells take up MUC2 and glycans associated with MUC2 inhibited dendritic cell inflammatory responses, but not tolerogenic responses, by inducing a galectin-3–Dectin-1–FcγRIIB receptor complex that activated β-catenin, resulting in nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) inhibition. Thus, mucus not only merely forms a nonspecific physical barrier, but also constrains the immunogenicity of gut antigens by delivering tolerogenic signals to mucosal dendritic cells.

CONCLUSION

The interplay between immunostimulatory and suppressive activities of intestinal dendritic cells is important for both the induction of an immune response and the maintenance of local immunostasis. The continuing identification of dendritic cell subsets within the human intestine and the elucidation of signaling mechanisms that govern mucosal dendritic cell activity are crucial to better understand the aberrant immune responses associated with a range of gastrointestinal disorders, including inflammatory bowel disease, irritable bowel syndrome and food allergy. In addition, the recent exciting discoveries describing the factors that mediate microbiome–host dialog may provide new candidate molecules for the deliberate modulation of dendritic cell activity.

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Conflicts of interest

E.S. and S.S. have no conflicts of interest.

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